

Variation in Bacteriohopanepolyol content of soils and sediments

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Background

Bacteriohopanepolyols (BHPs) are pentacyclic triterpenoids biosynthesised by many prokaryotes to perform a regulating and rigidifying function in membranes analogous to that of some sterols in eukaryotes. BHPs have been detected in a wide range of prokaryotes. Many BHP structures are known, differing in the number, position and nature of the functional groups on the side chain (see handout). In some cases these BHPs are unique to specific bacterial sources and these relationships can be used to identify the presence of bacterial processes in the soil (Table 1).

This Study

The Palace Leas pasture plots, Cockle Park Farm, Hebron, Northumberland, have undergone a consistent fertiliser treatment regime of over 100 years. The BHP signature of these plots was investigated using HPLC analysis to identify any differences due to the differing fertiliser treatments. Plot 2 – manure treatment, Plot 6 – No applied fertiliser
Plot 7 – Nitrogen fertiliser only Plot 8 – Phosphate treatment only
Plot 9 – Potassium fertiliser only.
The results are shown in Figs 1 and 2

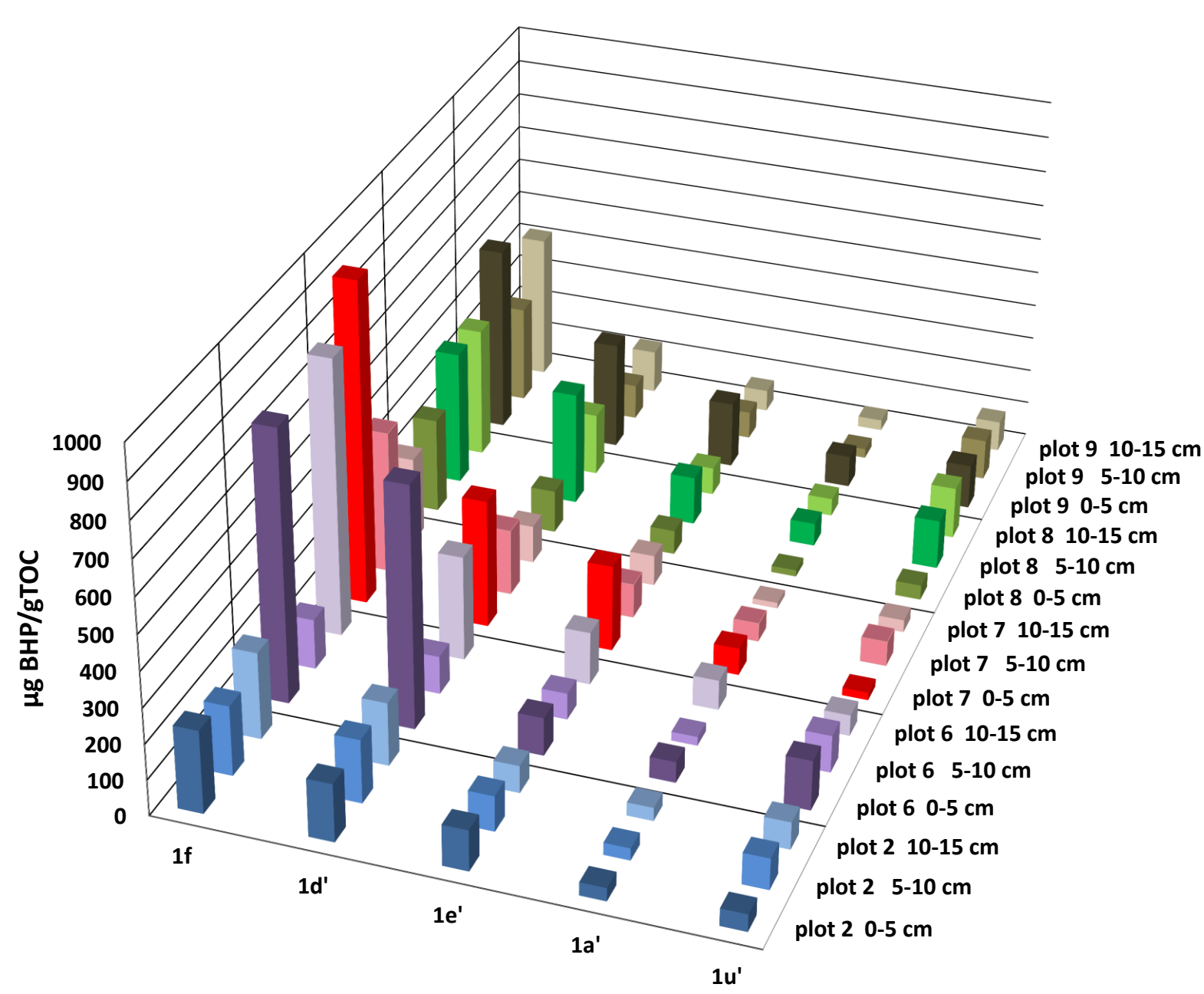


Fig. 1 Average Concentration of Major BHPs in Palace Leas Plots

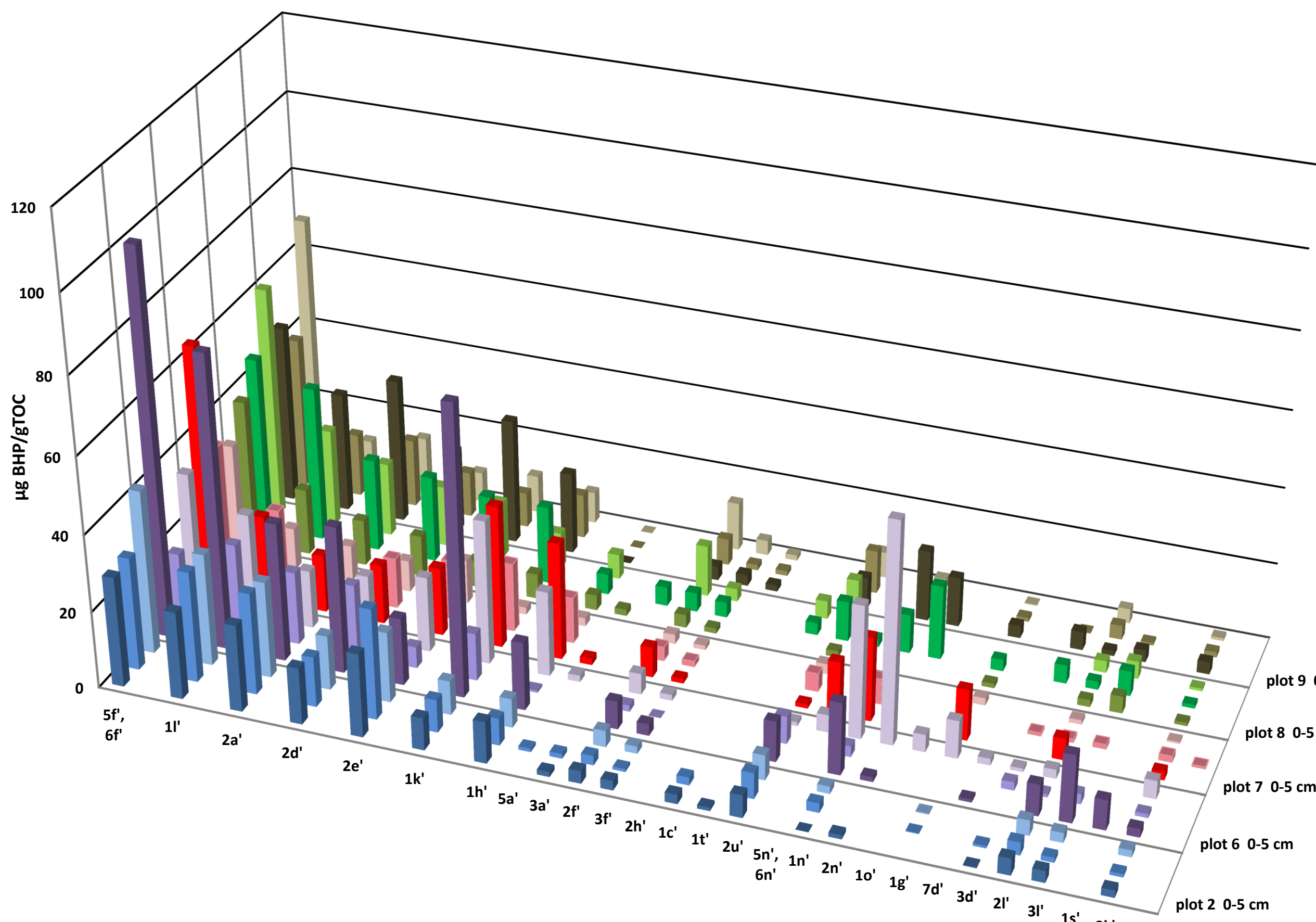


Fig. 2 Average Concentration of Minor BHPs in Palace Leas Plots

Results and Discussion

Up to 32 different BHPs were identified in the soil samples (Table 1).The BHPs can be split into major and minor components. The major components are 5 BHPs that dominate the total BHP concentration. Aminotriol (1f), BHT cyclitol ether (1d) and BHT (1a) are commonly found in all environments tested and adenosylhopane (1e) and adenosylhopane type-1 (1u) have been identified as soil marker BHPs, being unique to soils (Cooke et al, 2008). Potentially far more instructive are the minor BHPs that comprise the remaining BHP concentration. These BHPs indicate subtle differences between the various plots that may have arisen due to the various fertiliser regimes. For example plot 2 contains significant quantities of aminopentol (1c), a methanotroph marker (Table 1) whereas this BHP is absent from all other plots. This observation is in agreement with the expected increased presence of methanotrophs in manured plots (Seghers et al., 2003). Each plot displays significant differences in the presence / absence of specific minor BHPs or significant concentration differences indicating differences in the bacterial population. The differences can, however be hard to visualise so an alternative approach must be taken to highlight the impact of the BHP signatures. As previously stated many BHPs are strongly linked to specific bacteria and bacterial types (Table 1). Grouping the BHPs into bacterially related clusters enables PCA analysis of the results. Fig 3 shows the loadings plot for the analysis of the Palace Leas plots and clearly demonstrates that different bacterial groups have different influences on the plots. The scores plot (Fig 4) shows that the different plots are separated by the their BHP content. Again it is clear that for the case of plot 2 it is the presence of a methanotroph signature that distinguishes this plot. For plot 7 it is the diversity of the BHP signature that is important, indicating either a diverse bacterial population or conditions that require the production of a wide range of BHPs.

Table 1. Relationship between BHPs and Source Bacteria

Abbreviated name	Structure Number	Known Source organisms*
BHT	1a'	Various
Aminotriol	1f'	Various
BHT ce	1d'	Various
BHT glu	1g'	Various
BHpentol ce	1i'	various
Unsat. aminotriol	5f', 6f'	<i>Rhodopseudomonas palustris</i> ⁴⁷
Adenosylhopane	1e'	Purple non-sulfur bacteria ^{14,22,26,36,47} <i>Nitrosomonas europaea</i> ^{39,47} <i>Bradyrhizobium japonicum</i> ⁴⁶
2-me adenosylhopane	2e'	<i>Bradyrhizobium japonicum</i> ⁴⁶
Adenosylhopane type- 1 ^{1a}	1u'	Purple non-sulphur bacteria ⁴⁷
Δ ⁶ or 11'-BHT	5a', 6a'	Acetic acid bacteria ^{27,48}
3-me BHT	3a'	Acetic Acid Bacteria, ⁴⁸
3-me BHT ce	3d'	<i>Gluconacetobacter xylinus</i> ⁴⁸
3-me BHpentol ce	3i'	<i>Gluconacetobacter xylinus</i> ⁴⁸
2-me BHT	2a'	Cyanobacteria ^{3,13,40,44,50,55} <i>Rhodopseudomonas palustris</i> ³² <i>Methylobacterium organophilum</i> ²⁴
2-me aminotriol	2f'	Cyanobacteria ⁵⁶ <i>Rhodopseudomonas palustris</i> ³²
Unsat. BHT pentose	5n', 6n'	Cyanobacteria ⁵⁰
BHT pentose	1n'	Cyanobacteria ^{36,50}
2-me BHT pentose	2n'	Cyanobacteria ⁵⁰
2-me BHT ce	2d'	Cyanobacteria ⁵⁰
2-me BHpentol cyclitol ether	2i'	Unknown, potentially cyanobacteria as 2-Methyl
3-me aminotriol	3f'	Methanotrophs ³⁷
Aminotetrol	1h'	Methanotrophs ^{23,24,36,42,43,56} <i>Desulfotomaculum</i> sp. ⁴
2-me Aminotetrol	2h'	Methanotrophs
Aminopentol	1c'	Methanotrophs ^{11,23,36,42,43,56}
Guanidine sub. BHT ce	1s'	Methylotrophic bacteria ⁴⁷
Hopane lactone	1t'	Ammonia oxidising bacteria ^{39,47} PNSB ⁴⁷
Unsat. BHpentol ce	5i', 6i'	<i>Gluconacetobacter xylinus</i> ⁴⁸
2-me adenosylhopane type-1'	2u'	Unknown
BHpentol pentose	1o'	Unknown
Unsat. 2-me BHT ce	7d'	Unknown
BHpentol glu	1q'	Unknown
BHhexol ce	1k'	Unknown
2-me BHhexol ce	2k'	Unknown
3-me BHhexol ce	3k'	Unknown

* see Talbot et al., 2008 and subsequent references;

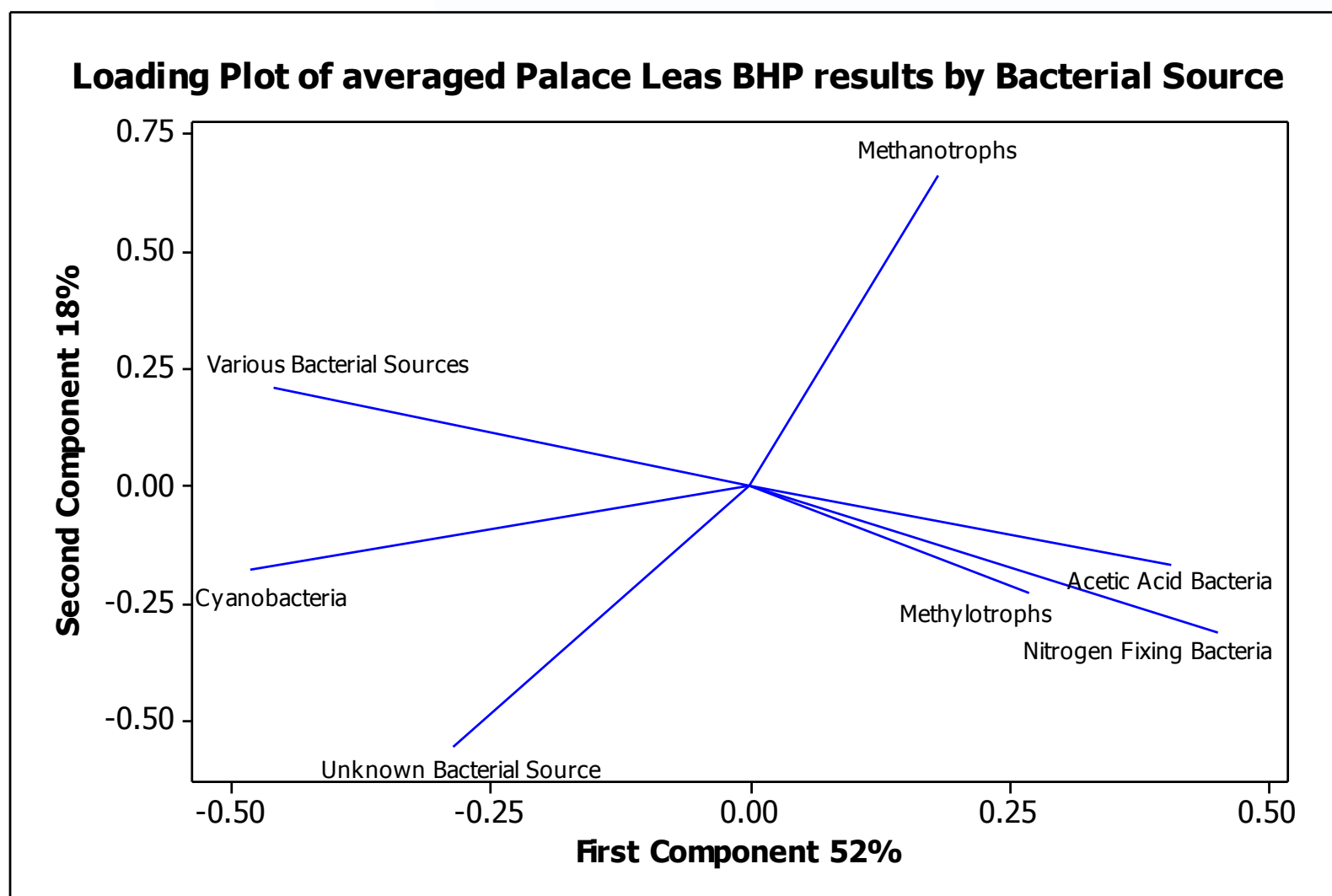


Fig. 3 Loading Plot of averaged Palace Leas BHP results by Bacterial Source

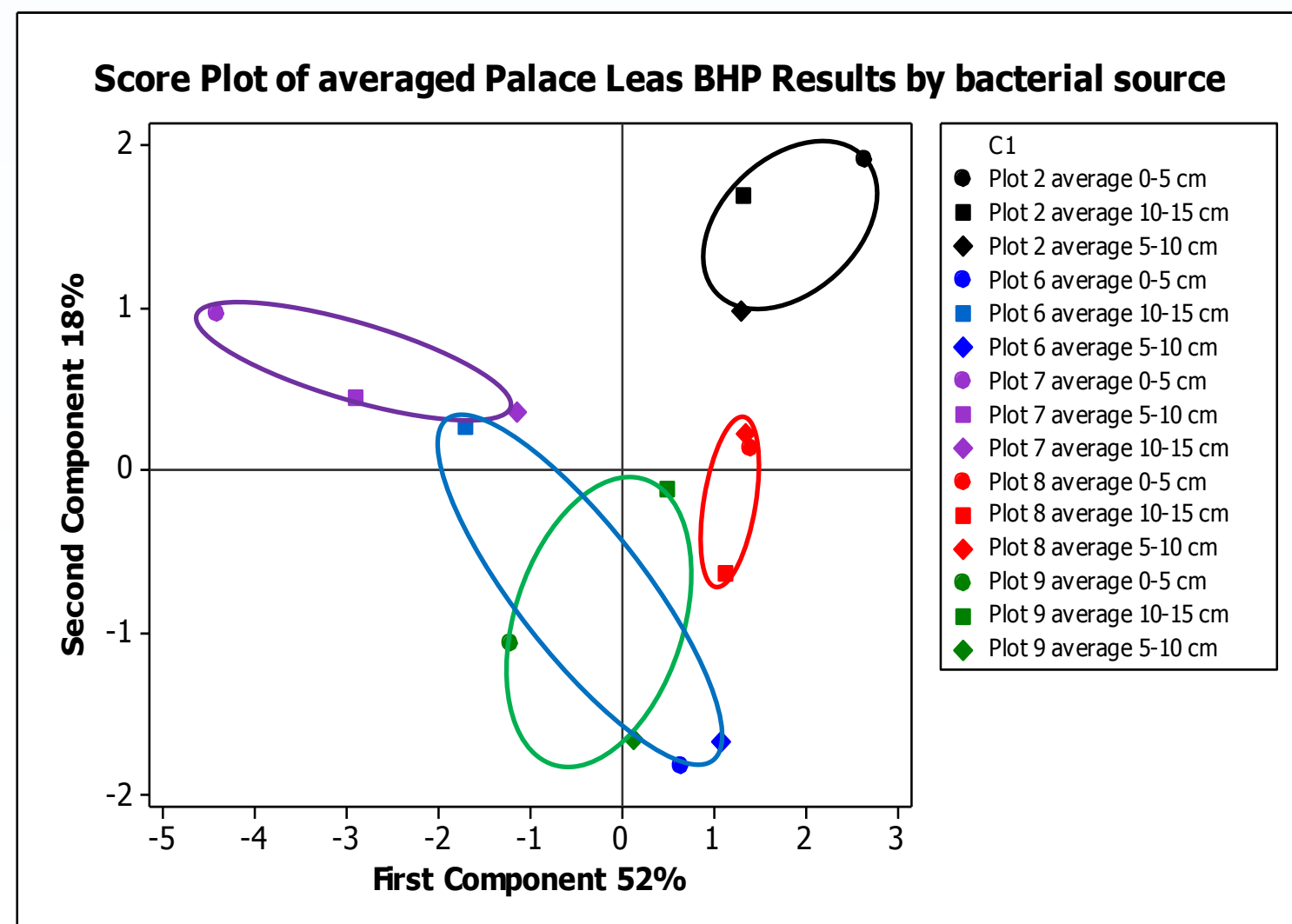


Fig. 4 Scores Plot of averaged Palace Leas BHP results by Bacterial Source

Global Relationships

An identical analysis was conducted on a suite of global soil and river fan sediments (Cooke 2010; Cooke et al., 2008a, 2008b, 2009; Redshaw et al., 2008; Xu et al., 2009), to assess the potential of this approach to separate soils and sediments. The loading plot (Fig. 5) indicates that again the presence of methanotrophs and the ability to identify a wide range of BHPs are significant factors in the separation of the soils and sediments (Fig. 6). There is a clear distinction between many of the locations. The majority of the soils are separated by the relationship between the common known sources and the dominant variously sourced BHPs. The presence of methanotrophs and the lack of soil markers separates the sediments from the soils. The most unusual result was the location of the Palace Leas plot soils and a set of soils from a NE woodland soil. These are distinct from the other soils and it is believed that the diverse BHP signature from these soils influences their position on the plot.

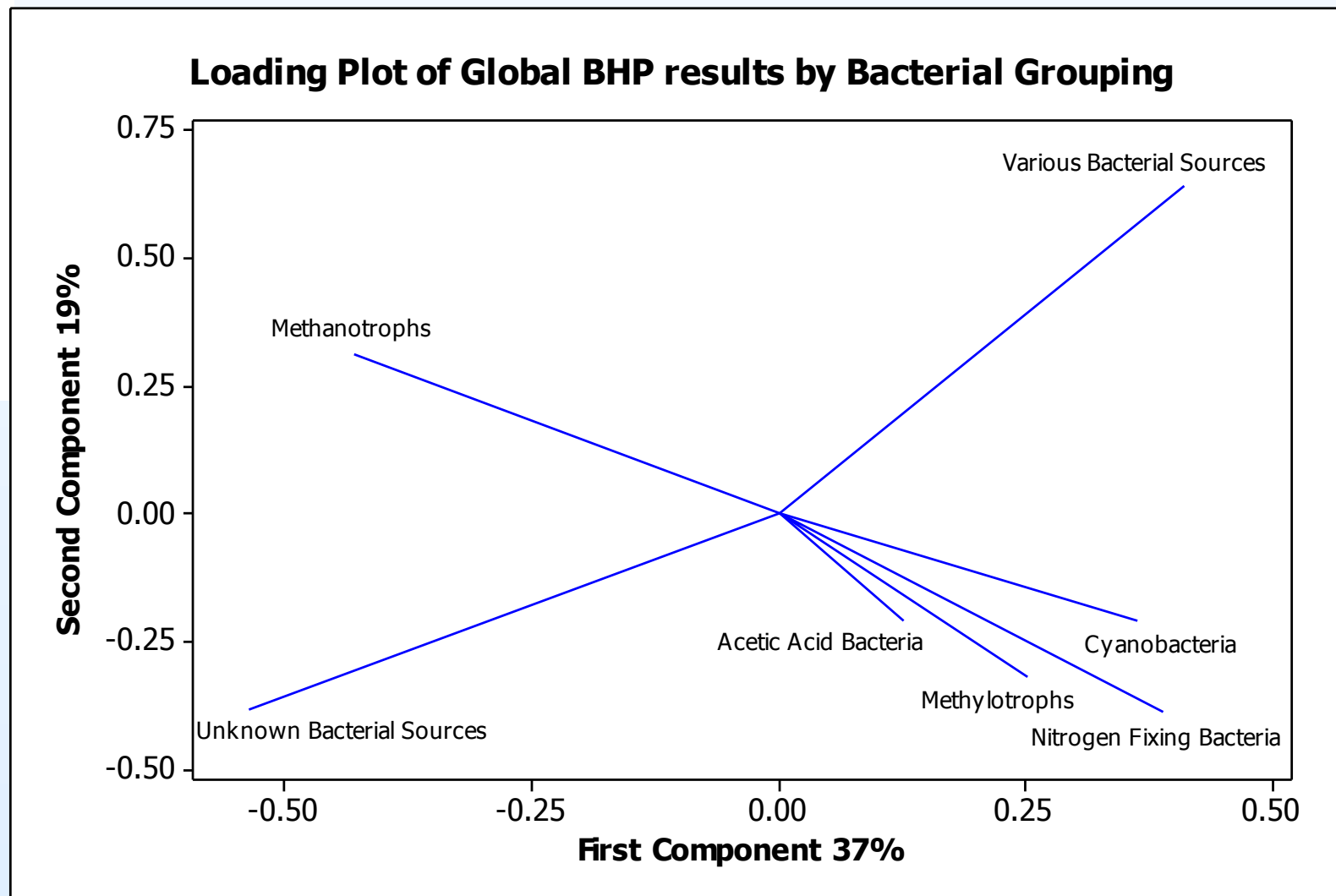


Fig. 5 Loading Plot of Global BHP results by Bacterial Grouping

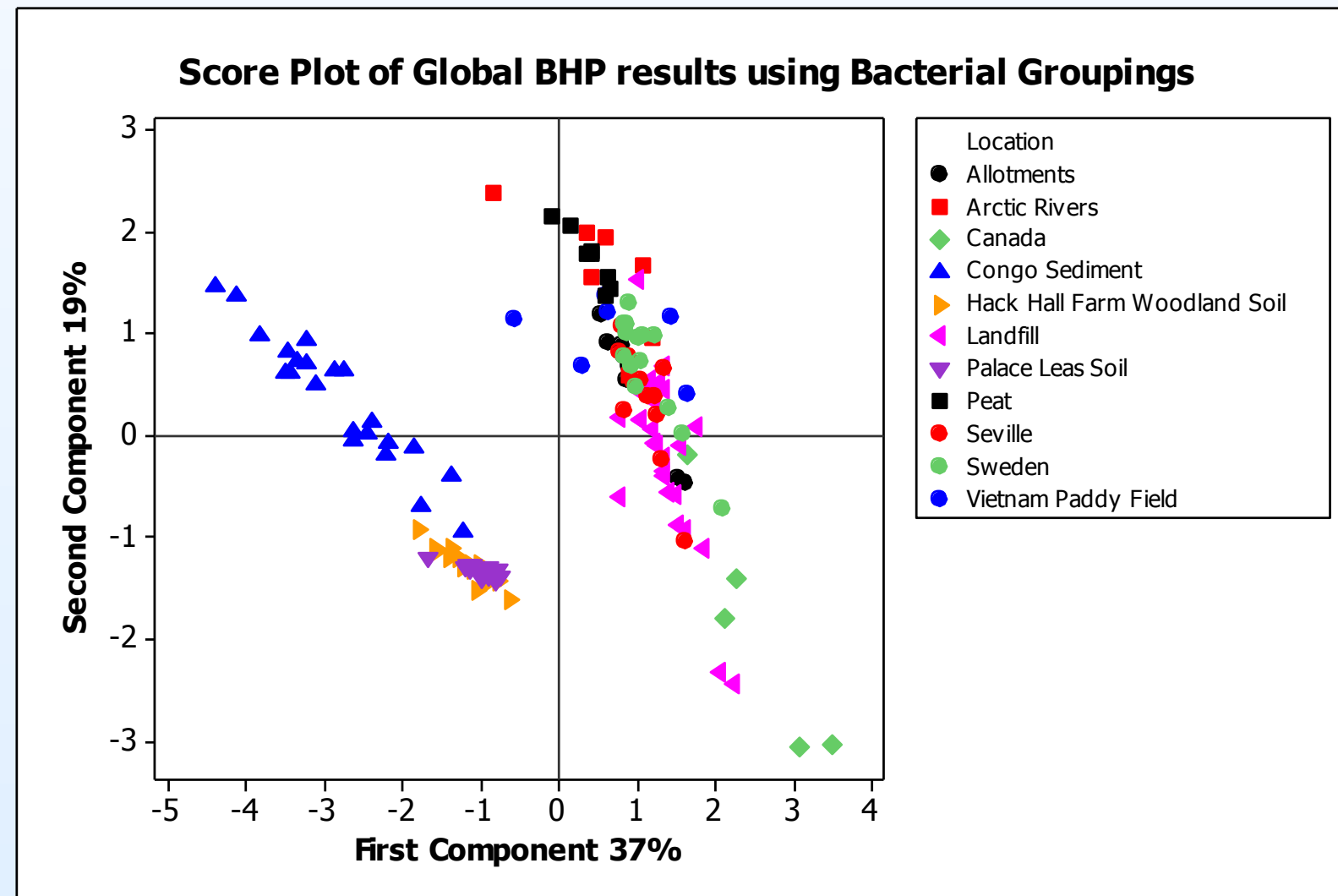


Fig. 6 Scores Plot of Global BHP results by Bacterial Grouping

Conclusions

It is clear from these results that there is great potential in the use of BHPs to identify soil bacterial processes and distinguish between different soils. However much work is needed to be conducted on a much greater suite of soils and on improving the relationship between the BHPs and the source bacteria, especially in those BHPs that are currently of unknown origin.

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